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FASTest® D4T bovine

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In vitro diagnosticum

Test-kit for the qualitative detection of Bovine Coronavirus (BCV), *Cryptosporidium parvum*, *E.coli*-K99 (F5) and Rotavirus in feces of cattle

INSTRUCTIONS FOR USE

Supplied Exclusively To The UK
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1. INFORMATION ON THE TEST-KIT

TEST-KIT COMPONENTS

1 test-kit **FASTest® D4T** bovine contains:

- 10 or 25 revolver test tubes **R** with 4 dipsticks each, coated with monoclonal antibodies against BCV, *C. parvum*, *E. coli*-K99 (F5) and Rotavirus
- 10 or 25 sample tubes **P** with 2.0 ml buffer diluent each
- 1 instructions for use

STABILITY AND STORAGE

Store at 15–25°C
Expiry date – see label

APPLICATION AND ABBREVIATIONS

- For veterinary use only
- LOT** Lot number
- In vitro diagnosticum
- Do not use test-kit components from different kits, lot numbers or beyond stated expiry date.
- Follow instructions for use precisely
- TL** – TEST line, **CL** – CONTROL line, **LF** – Lateral flow
- R** - Revolver test tube, **P** – sample tube

LIABILITY

The entire risk due to the performance of this product is assumed by the purchaser. The manufacturer shall not be liable for indirect, special or consequential damages of any kind resulting from the use of this product.

2. INTRODUCTION

Diarrhoea caused by Bovine Coronavirus (BCV), *Cryptosporidium parvum* (*C. parvum*), enterotoxigenic *Escherichia coli* K99 (F5) and Rotavirus (RV), is a major cause of severe illness and death in calf rearing. Especially *C. parvum*, but also Rotavirus are responsible for infectious diarrhoea in domestic animals and humans (cryptosporidiosis = zoonosis).

Clinical symptoms can vary due to age and immune state of the animal. Particularly in calves within the first two weeks of life, severe diarrhoea can occur (neonatal diarrhoea, ND), often with high mortality! Due to the high infection pressure, a general diarrhoea problem could occur in the farm. In this case, aetiological diagnostics of all bovines, regardless of age (asymptomatic shedders), and a check-up of feeding and keeping management is advisable.

Special attention should be paid to Cryptosporidiosis, because it is the only one that can be treated specifically. Here, an early diagnosis is a prerequisite for a successful therapy, because medication has to begin within 24 hours after onset of diarrhoea. Therefore, an early detection is necessary for a successful treatment as well as fast initiation of prophylactic measures.

ACCURACY

	Sensitivity	Specificity
BCV	100%	100%
<i>C. parvum</i>	100%	95.8%
<i>E. coli</i>	88.9%	100%
Rotavirus	96.4%	95.4%

(Comparison Method: ELISA)

3. INFORMATION ON THE SPECIMEN MATERIAL

Due to the normally inhomogeneous or nest-like dissemination of antigens in the feces, the specimen material has to be mixed up homogeneously (spatula, vortex-mixer) before sampling.

For the test, the required amount of feces as described in issue 4.b/Specimen collection and preparation, is needed. The amount depends on the consistency of the sample. Use the attached spoon.

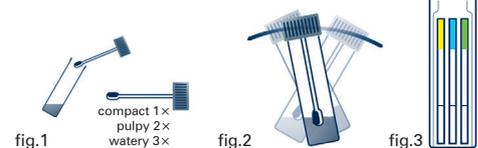
Non-cooled (15–25°C), the sample should be tested within 4 hours! At 2–8°C, the sample can be stored up to 4 days, permanently at minimum –20°C.

Keep in mind that the sample material, as well as all used test-kit components, should have reached room temperature at the time of application.

Endogenous and exogenous interfering substances of the sample (e.g. proteases, mucosa components, blood, but also viscosity, pH-value as well as grass and cat litter) can cause interferences (matrix effects) that can influence the target measurement. These can lead to an impaired LF and/or unspecific reactions on the TL and CL.

4. SPECIMEN COLLECTION AND PREPARATION

- Open the sample tube **P** with the buffer diluent.
- Mix the feces sample homogeneously (applicator, vortexer). Then mix the required sample

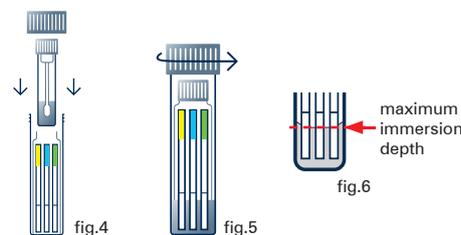


volume (fig.1: compact: 1 level spoon, pulpy: 2 level spoons, fluid-watery: 3 level spoons of feces) steadily into the buffer diluent.

- Close **P** tightly and rotate it easily to get the mixture as homogeneous as possible (fig.2). No sedimentation required. Immediately start with the test procedure 5.1.

5. TEST PROCEDURE

- Remove the revolver test tube **R** from the test-kit shortly before use.
- Open **R**, remove the red desiccant disk (fig.3) and introduce **P** containing the sample-buffer mixture (SBM) vertically into **R** (fig.4).
- Turn the cap of **R** until hearing a clicking noise twice (fig.5). The SBM of **P** will run into **R**. To ensure a proper LF, the liquid level must not exceed the white sucking pad of the dipstick (fig.6, see also 7. Precautions for users*).
- Place **R** on a flat and horizontal surface for incubation.



6. READING OF THE TEST RESULT

Read the test result after 10 minutes by turning **R** slowly around its own axis (fig.5). Positive test results may be observed earlier, depending on the concentration of antigen in the sample.

POSITIVE TEST RESULT (fig.7)

A pink-purple **TEST line of any intensity** (varying from very weak to strongly intensive) and a pink-purple **CONTROL line** appear.

NEGATIVE TEST RESULT (fig.8)

Only a pink-purple **CONTROL line** appears. This line indicates, irrespective of its intensity, that the test has been performed properly.

INVALID TEST RESULT

No **CONTROL line** visible. The test should be repeated using a new **P** and **R** *.

fig.7
POSITIVE TEST RESULT using the example of the Crypto Strip



fig.8
NEGATIVE TEST RESULT



7. PRECAUTIONS FOR USERS

- The guidelines for working in medical laboratories must be observed. It is recommended to wear disposable gloves and other personal protective equipment (protective clothing, possibly a face mask). Wash and disinfect hands after completing the test.
- Label sample material, associated sample tube **P** and revolver test tube **R** to ensure a precise assignment.
- Use a new sample tube **P** and revolver test tube **R** for each sample.
- The buffer diluent contains low concentrations of toxic sodium azide as a preservative, therefore avoid skin/eye contact and/or ingestion.
- The sample material must be seen as potentially infectious and disposed of accordingly, together with the used test-kit components.

* If the LF does not appear one minute after closing the cap of **R** completely, swing **R** carefully 1–2x in a circle.

If the LF still does not appear, maybe a too large amount of feces was used. The test must be completely repeated/rescheduled. Carefully observe the advice for sample preparation (also see 4.b/ Specimen collection and preparation and fig.1).

8. TEST PRINCIPLE

The **FASTest® D4T** bovine is based on latest rapid immunochromatographic technique. For each of the four pathogens of ND complex, there is an appropriate dipstick placed in the revolver test tube **R**. The dipsticks are characterised in colour as well as in writing: **Rota** = orange-red, **Corona** = yellow, **E.coli K99** = blue, **Crypto** = green.

The antigens in the feces sample will react at the conjugate pad with mobile monoclonal pathogen-specific antibodies (mAbs) bound to gold particles. Migrating ("lateral flow", **LF**) along the nitrocellulose membrane, these specific antigen-antibody complexes are bound by fixed mAbs producing a pink-purple **TEST LINE (TL)**. These mAbs guarantee a high level of specificity for the aetiological detection of the particular pathogen.

The intensity or width of the TL depends on the concentration of each antigen in the introduced amount of sample.

A correct test procedure will be indicated by a second, pink-purple **CONTROL line (CL)**.

In contrast to microscopic test methods depending on intact oocysts, the dipstick for *C. parvum* of **FASTest® D4T** bovine also detects surface antigens of vegetative *Cryptosporidium* forms or fragments of all *Cryptosporidium* forms, respectively.

Please note the two light-green quality control lines on **TL** and **CL**. They indicate that the test membrane is of good quality and can be used.

9. INFORMATION FOR THE INTERPRETATION

- The interpretation of the test result should always be based on anamnestic and clinical data as well as the therapy and prophylaxis possibilities.
- TL can vary both in intensity (from weak to intense pink-purple) and width. Therefore, any pink-purple line appearing within the required incubation time is to be interpreted as a positive test result.
- Any non-described colour or contour variation of TL and CL within the indicated incubation time or after more than 15 minutes (e.g. greyish, shadow-like lines) has to be considered as unspecific reaction and therefore as a negative test result.
- Because of intermittent antigen shedding, with ongoing diarrhoea a single negative test result should be confirmed by testing a **serial feces sample (individual testing of at least three consecutive feces samples)**.
- "Intensity of diarrhoea" can vary individually (age, immune status) or could not appear despite of a positive test result (asymptomatic eliminators!)
- Due to medical therapy, *C. parvum* surface antigens could be shed short-term and in a higher rate because of the additional shedding of vegetative *C. parvum* cyclus forms and cause positive test result despite of therapy for a short time.